Research Article

**Capparis spinosa: a comparative study of raw and processed fruits**

Mahnaz Khanavi¹,², Leila Ara³, Narjes Khavassí⁴, Homa Hajimehdipoor⁵,*

¹ Department of Pharmacognosy, Faculty of Pharmacy, Tehran University of Medical Sciences, Tehran, Iran  
² Faculty of Land and Food Systems, University of British Columbia, Vancouver, B.C. Canada  
³ Traditional Medicine and Materia Medica Research Center, Shahid Beheshti University of Medical Sciences, Tehran, Iran  
⁴ Metabolic Diseases Research Center, Zanjan University of Medical Sciences, Zanjan, Iran  
⁵ Traditional Medicine and Materia Medica Research Center and Department of Traditional Pharmacy, School of Traditional Medicine, Shahid Beheshti University of Medical Sciences, Tehran, Iran

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**ABSTRACT**

**Background:** *Capparis spinosa* is used in different countries for prevention and treatment of various diseases. Its fruits are usually used in processed form especially with vinegar. **Objective:** The aim of the present study was investigating the effect of processing on some phytochemical constituents and biological activities of *C. spinosa*. **Methods:** The fruits were processed with grape vinegar. Total phenolics and alkaloids contents of the raw and processed fruits were determined by Folin-Ciocalteu and titration methods, respectively. For quantification of rutin, quercetin and α-tocopherol, R-HPLC was used. Cytotoxic activity of the fruits was determined by MTT assay. Antioxidant properties were evaluated by DPPH and FRAP techniques. Fingerprinting of raw and processed fruits were prepared using TLC and evaluated by TLC densitometry instrument. **Results:** The results demonstrated that while total phenolics, total alkaloids and quercetin contents were decreased in the processed fruits, rutin concentration remained unchanged. Antioxidant activities of processed fruits increased using both DPPH and FRAP methods. Raw and processed fruits showed no cytotoxic effect on MCF-7, HepG-2 and MDBK cell lines up to the concentration of 100 µg/ml. The fingerprints of the fruits were different which admitted the change in the fruit constituents due to processing. **Conclusion:** It seems that processing with vinegar lessens the unpleasant taste of the plant due to alkaloids and increased the antioxidant effects; therefore, it would be more suitable for use in some diseases such as diabetes and hepatitis as it is used in folklore and traditional medicine.

**Abbreviations:** DPPH, 2,2-diphenyl-1-picrylhydrazyl; FRAP, ferric reducing ability of plasma; R-HPLC, reverse phase high performance liquid chromatography; TLC, thin layer chromatography; MDBK, madin-darby bovine kidney cells, HepG-2, hepatocellular carcinoma cells; MCF-7, michigan cancer foundation-7 breast cancer cells; MTT, 3-(4,5-Dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide; TPTZ, (2,4,6-tripyridyl-s-triazine); BHT, Butylated hydroxytoluene; ITM, Iranian traditional medicine

* Corresponding author: hajimehd@sbmu.ac.ir

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1. Introduction

Medicinal plants have been used as effective and almost safe therapeutic agents from ancient times. Many of them have multiple properties and use for diverse diseases. Moreover, they have shown fewer side effects than chemical drugs [1]. Nowadays, medicinal plants and their products are considered as an important source of bioactive constituents such as phenolics and flavonoids which are important in prevention and treatment of many diseases like cardiovascular and neurodegenerative diseases [2-4]. Capparis spinosa L., caper, (Capparidaceae) is one of the economical plants widely distributed in different region of the world [5] and is used in traditional medicine of many countries for diverse illnesses [1]. Several countries such as Greece, Italy, Spain and Turkey widely produce caper and export its different products to another countries which is used especially in food industry [6]. During the past two decades, more attention has been paid to phytochemistry and biological effects of C. spinosa. Phytochemical analysis demonstrated that the plant is rich in poly phenolics compounds which have been considered as the most responsible agents for health promoting effects of the species [7-9]. In addition, the plant contains alkaloids, glucosinolates, terpenoids and tocopherols [1]. During different studies, it has been proved that C. spinosa possesses various biological effects including antioxidant, anti-diabetes, antimicrobial, anticancer and hepatoprotective properties [10-15]. The fruits which are widely used, have bitter taste. Different methods are used to make fruits suitable for use such as fermentation [16] and pickle preparation. In Iran, especially in Iranian traditional medicine (ITM), fruits are used in pickle form for treatment of CNS and liver diseases [17]. Since, processing may change the plant constituents, in this research, the effect of vinegar processing on phenolics, alkaloids, rutin, quercetin and α-tocopherol contents, antioxidant and cytotoxic effects of caper have been investigated. Moreover, TLC fingerprint of raw and processed capers have been prepared.

2. Materials and Methods

2.1. Plant Material

Caper (ripped fruits) were collected in June 2016 from Pars Abad Moghan (Ardabil Province) and identified in Herbarium of Traditional Medicine and Materia Medica Research Center, Shahid Beheshti University of Medical Sciences, Tehran, Iran (voucher no. 3969 TMRC).

2.2. Chemicals

Folin Cio-calteu and pyrogallol were purchased from Merck Co. (Germany). Rutin, quercetin, α-tocopherol, DPPH, BHT and MTT were prepared from Sigma-Aldrich Co. (Germany). TPTZ was from Fluka Co. (UK) and grape vinegar was from Varda Co. (Iran). Thin layer chromatography was performed by using silica gel plates (Merck, Germany). All other solvents were in analytical grade (Merck, Germany). HepG-2, MCF-7 and MDBK cell lines were purchased from Pasteur Institute of Iran (Iran).

2.3. Pickle preparation

One thousand grams of intact fresh fruits was macerated in water (1:10) for 24 h. The mixture was sieved and again macerated in water for further 24 h. This process was repeated for 10 times and then the fruits were decocted in water for 1 min (three times after refreshing the water). Finally, the fruits were mixed with grape vinegar 30% and deposited for 30 days at room temperature. After that, the fruits separated from vinegar, dried in the shade and powdered.
2.4. Total phenolics contents

Total phenolics contents of caper were determined by Folin ciccalteu method by using pyrogallol as the standard material [18].

2.5. Rutin and quercetin contents

HPLC method was used for quantification of rutin and quercetin in the caper fruits [19].

**Test solution:** One gram of the powdered fruits was placed in a 5 ml conical flask and 4 ml methanol was added to the sample. The mixture was treated in an ultrasonic bath for 30 min and the volume adjusted with methanol to 5 ml. A portion of this solution was filtered through a membrane filter.

**Standard solution:** Quercetin and rutin (5 mg each one) were dissolved in methanol and diluted to 5 ml with the same solvent (stock solution). A diluted solution (5 ppm) was prepared from stock solution.

HPLC condition was as following: AQ C18, ACE, 4.6x250 mm, 5µm column, formic acid 1% in water: methanol with gradient mode (Table 1) as mobile phase, flow rate 0.6 ml/min in wave length of 257 nm, column temperature 25 °C, injection volume 100 µl.

2.6. Determination of total alkaloids as hyoscyamine

Total alkaloids of raw and processed fruits were determined according to Iranian Herbal Pharmacopoeia [20]. In this method, at first alkaloids were extracted by using mixture of ammonia/ethanol/diethyl ether, then the extract was acidified and alkaloids were transferred to aqueous phase. This solution was again alkalized by ammonia and extracted with chloroform. In the final process, chloroform fraction acidified and aqueous phase was titrated by NaOH. The percentage of total alkaloids was determined as hyoscyamine.

2.7. α-Tocopherol content

HPLC was used for α-Tocopherol determination in the fruits. Sample and standard material were dissolved in methanol and injected to HPLC with following condition: C_{18} AQ. 250x4.6 mm, 5µm column in 25°C, methanol:H₂O 96:4 as mobile phase, flow rate of 1.5 ml/min, 20µL injection volume in wave length of 284 nm. The content of α-Tocopherol in the sample was determined by comparing to the standard.

2.8. Fingerprinting of raw and processed caper

Fingerprinting of caper was performed by using silica gel plates and n-butanol: acetic acid: H₂O 40:1:10 as stationary and mobile phase, respectively. For preparation of test solution, 1 g of powdered fruits was added to 5 ml conical flask and methanol was added. Sulfuric acid 10% in methanol was used as reagent. The plates were scanned under 366 and 700 nm by Camag TLC scanner 3.

<table>
<thead>
<tr>
<th>Time (min)</th>
<th>Methanol (%)</th>
<th>Formic acid 1% (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>0</td>
<td>5</td>
<td>95</td>
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<tr>
<td>10</td>
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<td>55</td>
<td>98</td>
<td>2</td>
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<tr>
<td>60</td>
<td>5</td>
<td>95</td>
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</table>

Table 1. Gradient condition for separation of rutin and quercetin in *Capparis spinosa* fruits
2.9. Antioxidant effect of the caper by Ferric Reducing Ability of Plasma (FRAP) method

The antioxidant activity of raw and processed fruits was determined by FRAP method [21]. Briefly, the FRAP reagent contained 2.5 mL of a 10 mmol/L TPTZ solution in 40 mmol/L HCl plus 2.5 mL of 20 mmol/L FeCl₃.6H₂O and 25 mL of 0.3 mol/L acetate buffer (pH 3.6) which was prepared freshly and warmed at 37 °C. Alliquots of 40 μL of sample (methanolic extract) were mixed with 0.2 mL distilled water and 1.8 mL FRAP reagent and were incubated at 37 °C for 10 min. The absorbance of the reaction mixture was measured at 593 nm. Calibration curve of FeSO₄.7H₂O was plotted by using different concentrations. FRAP value was determined for each solution and expressed as mmol FeSO₄.7H₂O/100 g extract. Butylated hydroxytoluene (BHT) was used as positive control.

2.10. Determination of 2,2-Diphenyl-1-picryl hydrazyl (DPPH) radical scavenging assay

DPPH radical scavenging activity of samples was measured according to previous works [21, 22] as following: 100 μL of 100 μM DPPH methanol solution was added to 100 μL of various concentrations of the methanolic extracts. The mixture was shaken and left at room temperature for 30 min. Then, the absorbance of the solutions was measured at 517 nm by an ELISA reader and antioxidant activity was calculated. IC₅₀ was calculated from the plot of inhibition percentage against extract concentration. BHT was used as positive control.

<table>
<thead>
<tr>
<th>Table 2</th>
<th>The results of analysis of raw and processed Capparis spinosa fruits</th>
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</thead>
<tbody>
<tr>
<td>Test</td>
<td>Raw caper</td>
</tr>
<tr>
<td>------------------</td>
<td>------------</td>
</tr>
<tr>
<td>Total phenolics content as pyrogallol (%)</td>
<td>0.19±0.01</td>
</tr>
<tr>
<td>Rutin content (mg/100g fruits)</td>
<td>3.10±0.03</td>
</tr>
<tr>
<td>Quercetin content (mg/100g fruits)</td>
<td>1.66±0.01</td>
</tr>
<tr>
<td>Total alkaloids content (%)</td>
<td>1.87±0.01</td>
</tr>
<tr>
<td>α-Tocopherol content (mg/100g fruits)</td>
<td>0.0</td>
</tr>
<tr>
<td>FRAP value as mmol FeSO₄.7H₂O/100g extract</td>
<td>16.05±0.36</td>
</tr>
<tr>
<td>DPPH radical scavenging activity (IC₅₀, μg/ml)</td>
<td>556.66±26.73</td>
</tr>
<tr>
<td>Cytotoxic activity on HepG-2 (IC₅₀, μg/ml)</td>
<td>&gt;100 μg/ml</td>
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<tr>
<td>Cytotoxic activity on MCF-7 (IC₅₀, μg/ml)</td>
<td>&gt;100 μg/ml</td>
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<tr>
<td>Cytotoxic activity on MDBC (IC₅₀, μg/ml)</td>
<td>&gt;100 μg/ml</td>
</tr>
</tbody>
</table>

2.11. Evaluation of cytotoxic activity

2.11.1. Cell lines: The cell lines were obtained from the Pasteur Institute, Tehran, Iran: HepG-2 (human hepatocellular carcinoma), MCF-7 (human breast adenocarcinoma) and MDBC (bovine kidney cells). MCF-7 cells were cultured in DMEM medium with 5% FBS, while the other two cell lines were maintained in RPMI 1640 medium with 10% FBS to obtain the desired growth.

2.11.2. MTT assay: Assessing the viability of the cells was carried out in a micro culture tetrazolium/formazan assay (MTT assay). 96-well plates were used and HepG-2 cells 15 ×10³, MCF-7 cells 8×10³ and MDBC cells 10×10³ were seeded in each well. They were then incubated at 37°C. After 24 h the medium was replaced with fresh medium containing different concentrations of the methanolic extracts to be tested. After 72h exposure of cells at 37°C to each sample, the medium was replaced with fresh medium containing MTT, with a final concentration of 0.5 mg/mL. The cells were incubated for another 4 h in a humidified
atmosphere at 37°C, then the medium containing MTT was removed and the remaining formazan crystals were dissolved in DMSO. The absorbance was recorded at 570 nm with an ELISA reader (TECAN). Tamoxifen was used as positive control. The relative cell viability (%) related to control wells containing cells, cell culture medium and DMSO 1% was calculated by \([A] / [A]_\text{control} \times 100\).

Where \([A]\) samples is the absorbance of test sample and \([A]_\text{control}\) is the absorbance of wells containing cells + medium + DMSO, 1%. To calculate IC\(_{50}\) values, viability (%) versus concentrations was graphed using the Microsoft Excel program [23, 24].

3. Results

The results of the comparison between raw and processed caper have been demonstrated in Table 2. As it is obvious, total phenolics content was decreased during processing because of the vinegar effect on phenolic compounds and probably destroying them. Quantization of rutin and quercetin as two major flavonoids in the fruits demonstrated that rutin was unchanged but quercetin decreased which caused lower concentration of total phenolics in processed fruits (Fig. 1). No α-tocopherol was found in the fruits. During fruit processing, total alkaloids content were decreased about 70%, however, antioxidant activity have increased. Both raw and processed fruits showed no cytotoxicity on the studied cell lines up to 100 µg/ml.

![Fig. 1. HPLC chromatogram rutin and quercetin (A), raw caper (B) and processed caper (C)](image-url)

The chromatograms of raw and processed caper were completely different (Fig. 2). Indeed, in 700 nm, three brown spots were observed in lower half part of the raw fruits chromatogram but no spot was in this part of processed caper chromatogram. In both chromatograms, several spots were obvious in the upper parts of the silica gel plate with different intensities. In...
nm, there were blue-violet spots in middle parts of the chromatogram in processed caper but these spots were very weak in the raw fruit chromatogram.

4. Discussion

*Capparis spinosa* is used in traditional and folklore medicine of many countries [1]. It is consumed as pickle form. The findings of the present study demonstrated that vinegar has certainly destructive effect on some phenolic components which cause decreasing of total phenolics content but for determining the vinegar effect on specific constituents, quantitative determination of each component should be performed. In a study on caper collected from Tafresh, Iran, quercetin content was found 9.6 mg/g which is more than our study (1.66 mg/100g) [25]. While other studies demonstrated high concentration of α-tocopherol in caper [26], no α-tocopherol was found in the fruits in our study which demonstrate the effect of different climatic situation on the plant metabolites.

Alkaloids as secondary metabolites play different roles in plants and usually induce a bitter taste. It seems that one of the reasons for several washing of the fruits with water before adding vinegar is omitting the bitter taste. Raw caper is very bitter and many of water soluble alkaloids omit during processing and their concentrations decrease after processing. In fact, processing improves the caper taste. The vinegar effect on caper components were established in fingerprinting study as well with distinct differences between TLC chromatograms of raw and processed fruits.

No cytotoxic effect was seen on HepG-2, MCF-7 and MDBK cell lines up to 100µg/ml. Other investigations have shown cytotoxic effects of different parts of *C. spinosa* on various cell lines. Al-daraji *et al.* demonstrated toxic properties of *C. spinosa* leaves on Hep-2 and HeLa cells in concentration of 125-1000 µg/ml in dose-dependent manner [27]. It has proven that seeds contain 38 kDa protein which is similar to imidazole glycerol phosphate synthase and it inhibits MCF-7, HT-29 and HepG-2 proliferation [28]. In another study, the effect of different extracts of the plant fruits on Hep-2 and HeLa cells have been investigated. The results showed polyphenolics extract were cytotoxic in concentration of 10000 µg/ml. Other extracts showed less cytotoxicity [29]. It is obvious that caper is cytotoxic only in high concentrations and in the recent study it established no toxic effect up to 100µg/ml.
which is common concentration for considering an extract to be toxic.

The results of antioxidant activity demonstrated that vinegar induced more radical scavenging and antioxidant activity. Different investigations have been performed on vinegars such as apple and grape vinegars and high antioxidant activity was established [30, 31]. In the other hand, vinegar has antioxidant activity which causes more antioxidant effect in processed caper. In a research on antioxidant properties of grape and its processed products, it has been proved that high effects of wine and vinegar rather than juice [30]. Regarding role of antioxidant compounds in prevention and treatment of many diseases such as cancer, hepatitis, diabetes, Alzheimer, Parkinson it may be concluded that the reason of usage of processed caper is its high antioxidant effect. Although better taste of processed caper is other reason for its processing.

5. Conclusion

It seems that processing with vinegar changes some components in the caper as it is obvious in finger printing study. Total phenolics and among them quercetin decreased during processing but antioxidant effect increased significantly due to vinegar constituents. Moreover, processing ameliorated the unpleasant taste of the plant through decreasing alkaloids; It should be noticed that the raw and processed caper showed no toxicity effect on normal cell line (MDBK); therefore, it could be consider safe for use. In general, regarding the mentioned changes in chemical constituents and biological activities of caper after processing, it would be more suitable for use in some diseases such as diabetes, hepatitis and neurodegenerative disorders as it is used in folklore and traditional medicine.

Author contributions

Homa Hajimehdipoor and Mahnaz Khanavi designed and supervised the study. Narjes Khavavisi prepared processed fruits and Leila Ara has performed experimental part.

Conflict of interest

The authors declare that there is no conflict of interest. The authors alone are responsible for the accuracy and integrity of the paper content.

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کبر: مقایسه میوه‌های خام و تدبیر شده
مهناز خانوی ۲، لیلا آرا ۳، نرجس خواصی ۴، هما حاجی مهدی پور ۵*
گروه فارماکوگنوزی، دانشکده داروسازی، دانشگاه علوم پزشکی تهران، تهران، ایران
۱دانشکده سیستم‌های خاک و غذا، دانشگاه بریتیش کلمبیا، ونکوور، کانادا
۲مرکز تحقیقات طب سنتی و مفراده بریتیش، دانشگاه علوم پزشکی زنجان، زنجان، ایران
۳مرکز تحقیقات بیماری‌های متابولیک، دانشگاه علوم پزشکی زنجان، زنجان، ایران
۴مرکز تحقیقات طب سنتی و مردمان، گروه داروسازی سنتی، دانشگاه علوم پزشکی شهید بهشتی، بهشتی، تهران، ایران

چکیده
مقدنه: میوه گیاه کبر در کشورهای مختلف برای پیشگیری و درمان بسیاری از بیماری‌ها مصرف می‌شود. این میوه‌ها معمولاً به صورت تدبیر شده با سرکه مورد استفاده قرار می‌گیرند. هدف از این مطالعه بررسی تاثیر تدبیر با سرکه بر بعضی از ترکیبات میوه گیاه و نیز برخی خواص پیوسته‌کننده و فیتوتراتوکسیک در میوه‌ها می‌باشد.

روش بررسی:
mیوه‌های گیاه بکر به صورت ترشی با استفاده از سرکه انگوری آماده‌سازی شدند. میزان ترکیبات فنلی و آلکالوئیدی‌های تام نمونه‌های خام و تدبیر شده با استفاده از روش فولین سیوکالتو و تیتراسیون اندازه‌گیری شدند. مقدار ترکیبات روتوئید با استفاده از HPLC تعیین شد. اثرات سایتوتوکسیک و مایاک سیسکو توسط روش MTT ارزیابی گردید. خواص آنتی‌اکسیدانی میوه‌های خام و مدبر با روش DPPH و FRAP تعیین شدند.

نتایج: نتایج نشان داد که میزان ترکیبات فنلی و آلکالوئیدی‌های تام نمونه‌های تدبیر شده کاهش یافته ولی میزان ترکیبات روتوئید به ترتیب با استفاده از HPLC و TLC تأیید گردید. خواص آنتی‌اکسیدانی میوه‌های خام و مدبر با روش DPPH و FRAP تعیین شدند. درمان سپارایزی از این میوه‌ها می‌تواند انتخاب مناسبی در درمان بیماری‌های متنوعی مانند دیابت و هپاتیت با توجه به مصرف آن در طب سنتی باشد.

اطلاعات مقاله
گزارشگر: کبر
تدریب: تدبیر
سرکه

کلمات کلیدی:
کبر
تدبیر
سرکه

تعیین دهنده تغییرات با استفاده از روش دیپتیم ۱۷ میکروگرم در میلی‌لیتر پودر. الگه‌سازی دو نمونه نژاد که شامل تغییرات ترکیبات ناشی از عمل تدبیر است. ترجیح‌گیری: به نظر من رشد تدبیر با سرکه طعم‌دهنده از آلکالوئیدی‌ها را کاهش داده و اثرات آنتی‌اکسیدانی را افزایش می‌دهد. بنابراین میوه‌های مدبر می‌توانند استفاده مناسبی در درمان بیماری‌های متنوعی مانند دیابت و هپاتیت با توجه به مصرف آن در طب سنتی باشند.

مخفف‌ها: هPLC (reverse phase, R-HPLC) reversal phase high performance liquid chromatography; (TLC) thin layer chromatography; (MDBK) madin-darby bovine kidney cells, (HepG-2) hepaticcellular carcinoma cells; (MCF-7) Michigan cancer foundation-7 (breast cancer cells); (MTT) 3-(4,5-Dimethylthiazol-2-yl)-2,5-diphenyldetrazolium bromide; (TPTZ) (2,4,6-tripyridyl-s-triazone); (BHT) Butylated hydroxytoluene; (ITM) Iranian traditional medicine

نویسنده مسئول: hajimehd@sbmu.ac.ir

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