

## Phytochemical Diversity of *Eremostachys laciniata* Bunge Populations in Iran

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### Abstract

**Background:** *Eremostachys laciniata* is known as "Chelle-Daghi" in Iran and its rhizomes are used as an emollient to relieve rheumatoid arthritis.

**Objective:** A phytochemical study was performed on 15 populations of *Eremostachys laciniata* to qualify the phytochemical variations.

**Methods:** Plants collected from their natural habitats and dried rhizomes of *E. laciniata* were milled and then Soxhlet-extracted by MeOH, and then total phenols were determined calorimetrically by the Folin-ciocalteau reagent and iridoid glycosides were measured by TLC method.

**Results:** The results of MeOH extracts showed that the highest amount with 14.85 gr/plant was obtained in the Areshtanab population. The populations had a significant effect on total phenolic concentrations ( $P < 0.001$ ) and the Areshtanab with 0.281 mg GAE/g DW had higher phenolic levels than other populations. The results of TLC of iridoids showed that 15 populations had about 11 iridoids components. Cluster analysis by using Ward's method detected that the studied populations of *E. laciniata* were separated into three different groups.

**Conclusion:** In summary, higher values of the total phenols and iridoids in the Areshtanab, Malek-Kian, and Til populations were considered to indicate higher levels of phytochemical heterogeneity and significant diversity between populations, which could be used to domesticate and develop the breeding programs.

**Keywords:** *Eremostachys laciniata* Bunge, Iridoids, Phytochemical diversity, Total phenol



## Introduction

The growing trend of increase in world population suggests that the world population in the first half of 21 century will exceed 10 billion people. Therefore, beside the food security, the efforts to provide health care will be one of the most important problems and concerns facing the international community [1, 2]. Plants are a rich source of secondary metabolites that have medicinal and aromatic properties. According to some estimates, at least 100,000 such secondary metabolites are now known to occur in 50,000 plant species and 4,000 new secondary metabolites are being discovered every year from a variety of plant species [3].

*Eremostachys laciniata* (L) Bunge (family: Lamiaceae alt. Labiatae; sub-family: Lamioideae) is a perennial herb with thick roots and pale-purple or white flowers. It is one of 15 species of *Eremostachys* grown in Iran and in other countries in the Middle-East, Western Asia, and the Caucasus [4, 5]. This genus has 60 species worldwide and five endemic species in Iran including *E. pulvinaris*, *E. codonocalyx*, *E. azerbaijanica*, *E. adenantha*, and *E. hyoscyamoides* [5].

In traditional medicine, the *E. laciniata* species is known in Iran as "Chelle-Daghi" and its rhizomes are used as an emollient to relieve rheumatoid arthritis [6]. Moreover, a decoction of the rhizomes and flowers of *E. laciniata* are traditionally used to treat allergies, headaches, and liver disease [7]. Antioxidant [8], antibacterial [9], antidepressant [10], anti-inflammatory [11] and analgesic [12] properties have been documented for *E. laciniata*.

Previous phytochemical study on *Eremostachys* genus established the presence of alkaloids, coumarins, flavonoids [13] and mono- and diterpene glycosides [14, 15], two ferulic acid derivatives [16], three phenylethanoid glycosides [17] and highly oxygenated flavonoid named loasifolin [18]. Phytochemical investigations have also reported the isolation of iridoids, phenylethanoid, and flavonoid derivatives from the aerial parts [19] and some iridoid and furanolabdane diterpene glycosides from the rhizomes of this medicinal species. Phytochemical studies also revealed the presence of various monoterpenes and sesquiterpenes among its essential oils [20].

Iridoids, the main secondary metabolites of this species, are secondary metabolites of terrestrial and marine flora and fauna and are found in a large number of plant families usually as glycosides. Structurally, they are cyclopentano pyran monoterpenoids and biogenetically and chemotaxonomically they provide a structural link between terpenes and alkaloids [21, 22]. Iridoids are present in a number of folk medicinal plants used as bitter tonics, sedatives, antipyretics, cough medicines, remedies for wounds, and skin disorders and as hypotensives. This fact encouraged to investigate the bioactivities of these phytochemicals. Intensive study of their bioactivity revealed that these compounds exhibit a wide range of bioactivity: cardiovascular, antihepatotoxic, choleric, hypoglycemic and hypolipidemic, antiinflammatory, antispasmodic, antitumor, antiviral, immunomodulator and purgative activities [22, 23, 24, 25]. Chemotaxonomically,

they are useful as markers of several genus in various plant families, such as, aucubin of *Plantago* (Plantaginaceae) asperuloside of *Galium* (Rubiaceae) and aucuboside (aucubin) and harpagide of *Scrophularia* (Scrophulariaceae) [23].

The aims of this study were: (i) quantifying phytochemical variation among 15 Iranian populations of *E. laciniata*, using TLC and total phenol analysis; and (ii) establishing phytochemical relationship among the populations.

## Materials and Methods

**Plant material:** The rhizomes of 15 populations of *Eremostachys laciniata* (L) Bunge, were collected from different parts of Iran in 2013-2014 (Table 1). The plants were identified by Flora Iranica [26]. A voucher specimen (TUM-ADE 0204) for this collection has been retained in the herbarium of the School of Pharmacy, Tabriz University of Medical science, Iran.

**Extraction and isolation:** The dried and ground rhizomes of *E. laciniata* (100 g) were milled and then Soxhlet-extracted, successively, with dichloromethane (DCM) and MeOH. The DCM extract was discarded and MeOH extract was concentrated by using a rotary evaporator at a maximum temperature of 45 °C [6].

**Total Phenol:** Total phenols in *E. laciniata* MeOH extract were determined calorimetrically at 725 nm (Human Crop, Xma-2000) with the Folin-ciocalteau reagent as previously done by Gutfinger [27].

**Thin layer chromatography (TLC):** Thin layer chromatography of iridoid glycosides containing fraction was performed using silica gel plate 60 F<sub>254</sub> (Merck, Germany). The solvent system was chloroform- methanol-water (60:40:4), where the sprayed reagent was anisaldehyde-sulphuric acid, followed by heating at 110°C for 5 min [28].

**Table 1- Collection Sites for *Eremostachys laciniata***

Site	Altitude (m asl <sup>+</sup> )	Latitude (N)	Longitude (E)	Mean annual temp. (°C)	Mean annual rainfall (mm)
Ajabshir	1,387	37° 40'	45° 51'	15.1	236.4
Areshtanab	1,967	37° 54'	46° 42'	13.5	222.5
Chelle-Khaneh	1,365	38° 15'	46° 01'	14.5	245.1
Heydarabad	1,449	38° 17'	45° 24'	16.2	341.8
Ilkhchi	1,426	37° 55'	46° 00'	15.1	236.4
Iranagh	1,986	37° 55'	46° 35'	13.5	222.5
Kaleybar	1,617	38° 45'	46° 58'	13.6	367.2
Malek-Kian	1,607	38° 03'	46° 32'	12.5	299.7
Marand	1,468	38° 24'	45° 44'	13.2	444.3
Meshanagh	1,634	38° 13'	45° 34'	16.2	341.8
Mianeh	1,811	37° 36'	48° 04'	15.3	337.4
Sad-Ammand	1,369	38° 11'	46° 08'	14.5	245.1
Sarab	2,122	38° 02'	47° 41'	10.2	276.4
Til	1,686	38° 16'	45° 29'	16.2	341.8
Zglojeh	1,949	37° 48'	46° 54'	10.5	421.5

<sup>+</sup> m asl: Meter after sea level



All data were subjected to statistical analysis (one-way ANOVA) by using MSTAT-C software. Differences between the treatments were performed by Duncan's Multiple Range Test (DMRT) at 1% probability level. The Ward's method was used for cluster analysis [29].

## Results

**Total phenolic compounds:** The results of MeOH extracts of roots of *E. laciniata* showed

that the highest amount of extract was in the Areshtanab population (Table 3).

Average total phenolic concentrations for 15 *Eremostachys* populations were determined by the Folin–ciocalteu assay and are presented in table 3. Populations had a statistically significant effect on total phenolic concentrations ( $P < 0.001$ ) by ANOVA analysis (Table 2). Mean comparison with DMRT showed that Areshtanab (0.281 mg GAE/g DW) had higher phenolic levels than other populations.

**Table 2- ANOVA of total phenols in 15 studied populations**

Source of variation	Degrees of freedom	Mean of squares (Total phenols)
Populations	14	0.0192 **
Error	30	0.00002
CV%		2.46

\*\* -Significant at 1% probability level

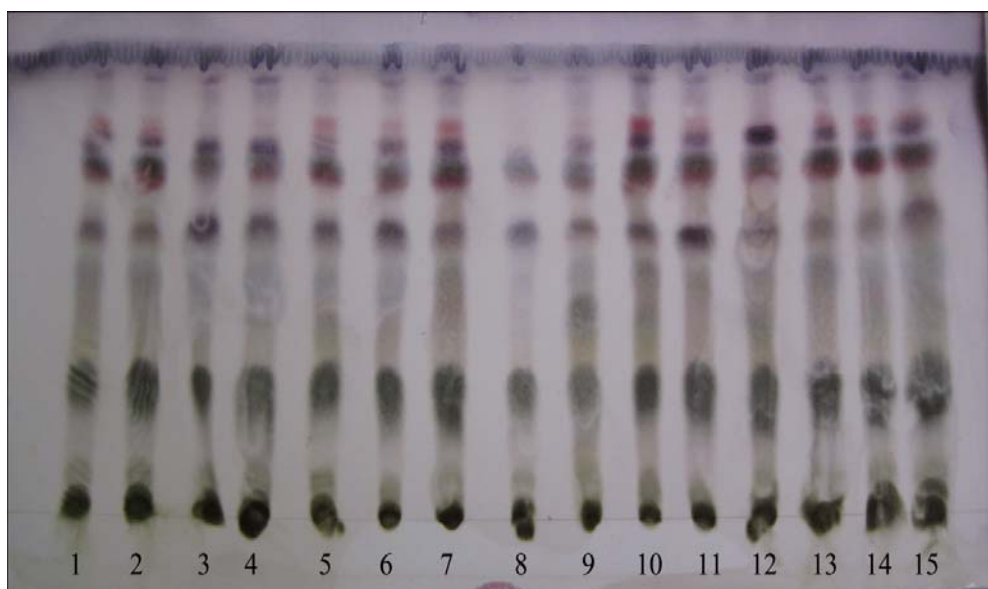
**Table 3- Mean comparison of extract and total phenols in 15 studied populations**

Populations	Extract in 100 g plant sample (gr)	Phenols in 1 gr extract (mg GAE/g DW)	Total phenols in 100 gr plant sample (mg GAE/g DW)
Ajabshir	5.31	0.012	0.065 i
Areshtanab	14.85	0.019	0.281 a
Chelle-Khane	13.70	0.016	0.220 d
Heydarabad	2.87	0.014	0.041 j
Ilkhchi	6.97	0.010	0.068 i
Iranagh	7.69	0.008	0.060 i
Kaleybar	8.85	0.016	0.139 g
Malek-Kian	13.35	0.019	0.258 b
Marand	12.73	0.020	0.251 b
Meshanagh	8.45	0.024	0.204 e
Mianeh	9.65	0.017	0.163 f
Sad-Ammand	11.82	0.013	0.158 f
Sarab	10.21	0.021	0.213 de
Til	12.48	0.019	0.236 c
Zglojeh	10.63	0.012	0.128 h

Means with the same letter(s) aren't significantly different at  $P \leq 0.01$  by DMRT

**Iridoids:** The results of TLC of iridoids (Figure 1 and Table 4) showed that, corresponding to  $R_f$ , 15 populations had about 11 iridoids components. The Malek-Kian, Mianeh, Chelle-Khane, Sarab, Meshanagh, Til, and Kaleybar showed all 11 iridoids

components but other population had not all components. The Iranagh had the lowest components. Wagner and Bladt [28] explained that iridoids color in the TLC was brown (light to dark), Blue, Purple, and gray and results of this study showed the same color (Figure 1).



**Figure 1-** Columns of iridoids TLC. 1, Malek-Kian; 2, Mianeh; 3, Zglojeh; 4, Ilkhchi; 5, Chelle-Khane; 6, Heydarabad; 7, Sarab; 8, Iranagh; 9, Sad-Ammand; 10, Meshanagh; 11, Areshtanab; 12, Ajabshir; 13, Til; 14, Kaleybar; 15, Marand

**Table 3-** Color, presence and quantity of the spots on the TLC chromatogram of *Eremostachys laciniata* roots

Color	Rf	Gray	Black	Purple	Gray	Brown	Gray	Purple	Brown	Blue	Purple	Blue
		0.01	0.30	0.46	0.53	0.61	0.66	0.74	0.76	0.8	0.85	0.95
Malek-Kian		1	2	1	1	1	1	1	2	2	1	1
Mianeh		1	2	1	1	1	1	1	2	2	1	2
Zglojeh		1	2	0	1	2	1	0	1	2	0	2
Ilkhchi		1	2	0	1	2	1	1	1	2	1	2
Chelle-Khane		1	2	1	2	2	1	1	1	1	1	1
Heydarabad		1	2	0	2	2	1	1	1	2	1	2
Sarab		1	2	1	2	2	2	1	2	2	1	2
Iranagh		1	1	0	0	1	0	0	1	0	0	1
Sad-Ammand		1	1	0	2	1	1	1	1	1	0	1
Meshanagh		1	2	1	2	2	2	1	2	2	2	2
Areshtanab		1	2	1	1	2	2	1	2	1	1	1
Ajabshir		1	2	0	2	1	2	1	2	2	0	2
Til		1	2	1	1	1	2	1	2	2	1	2
Kaleybar		1	2	1	1	1	1	1	2	1	1	1
Marand		1	2	0	1	1	1	1	2	1	1	1

**Cluster analysis:** A dendrogram was obtained from the cluster analysis by using Ward's method. This method detected that the studied populations of *E. laciniata* were separated into three different groups (Figure 2). The Til, Areshtanab, Malek-Kian, Chelle-Khaneh, and Marand were placed in the first group. This group had the higher levels of iridoids and total phenols. The second group consisted of the Zglojeh, Iranagh, Sad-Ammand, Mianeh, Sarab, and Meshanagh

populations. The Ilkhchi, Ajabshir, and Heydarabad populations were placed in the third group.

Results showed that among the populations, the Areshtanab, Malek-Kian, and Til populations can be distinguished from other populations by some phytochemical features that agreed with morphological [36] and cytological diversities [38], which could be used for domestication and development of breeding programs.

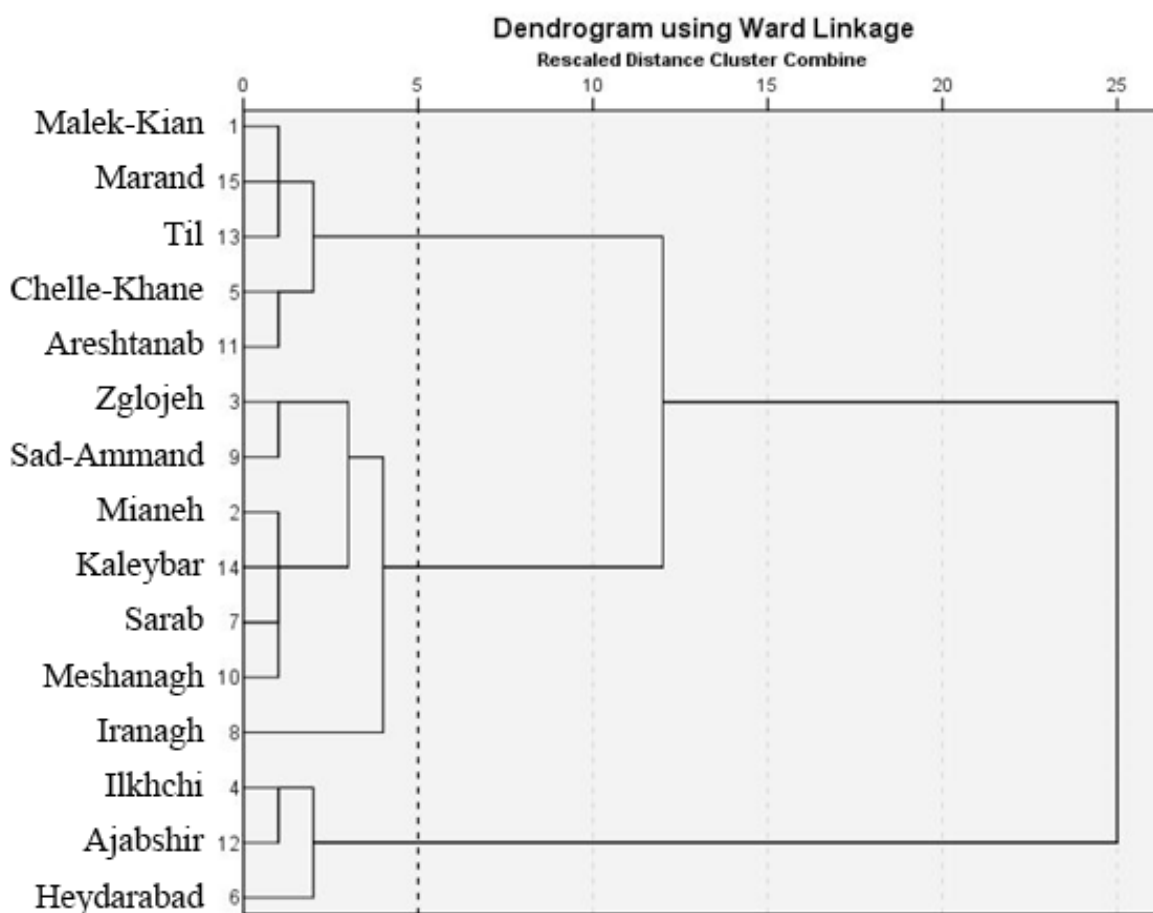


Figure 2- A dendrogram of cluster analysis by Ward's method

## Discussion

Cultivar is known to impact phenolic concentrations in a variety of plants, and the effect of genotype has been particularly well studied in fruits and vegetables [30, 31, 32]. The effect of cultivar on the phenolic composition and antioxidant properties of 15 different basil varieties was determined by Folin-ciocalteau reagent. Cultivar had a statistically significant effect on total phenolic levels and anthocyanin concentrations that could be used in breeding programs [33]. The phytochemical study of *Astragalus* showed existence of some secondary metabolites including phenols and flavonoids in this species. Therefore, total phenols were determined by Folin-ciocalteau reagent [34]. In the *Stachys* genus, the results of qualitative study of phytochemical compounds showed the presence of secondary compounds such as tannin, saponin, flavonoid, alkaloid, terpenoid, glycoside and steroid. In the quantitative study obtained the average amounts of total phenol  $74.56 \pm 4.343$  mg GAE g<sup>-1</sup> DW [35].

Morphological study of these 15 populations showed that the Til, Areshtanab and Malek-Kian populations had higher root

yield with 220, 171, and 164 gr/plant, respectively [36]. Therefore, considering the extract amount and root yield, it can be concluded that the Til, Areshtanab, and Malek-Kian populations had higher root and extract yield and iridoids and were usable in breeding programs.

Phytochemical investigation of a plant known as Tashnehdary (*Onosma chlorotricum*) was performed by TLC method and components R<sub>f</sub> was measured. The results confirms the antioxidant activity of some components in the lipophilic extract of *Onosma chlorotricum* root and consequently may confirm its effectiveness in some processes such as wound healing which antioxidant reactions are helpful [37].

## Conclusion

In summary, higher values of the total phenols and iridoids in the Areshtanab, Malek-Kian, and Til populations were considered to indicate the higher levels of phytochemical heterogeneity and significant diversity between populations, which could be used for domestication and development of breeding programs for this species.

## References

1. Craker LE and Gardner ZE. Medicinal plants and tomorrow's pharmacy. In R.J. Bogers, L.E. Craker, and D. Lange (eds.), Medicinal and aromatic plants. Proc. Frontis Workshop on Medicinal and Aromatic Plants, Wageningen, Nucleus for Strategic Expertise Wageningen University and Research Centre, Wageningen, The Netherlands. 2005, 29-41.
2. Farnia F and Jahangiri A. Phytopharmaceutical technology. Jahangiri press, Tehran, Iran. 1994, 186 pp.
3. Kumar J and Kumar Gupta P. Molecular approaches for improvement of medicinal and aromatic plants. *Plant Biotechnology Reports* 2008; 2: 93-112.
4. GRIN. Germplasm Resources Information

- Network. USDA - ARS, National Genetic Resources Program, Beltsville, MA, USA. 2013. National Germplasm Resources Laboratory, Available at: <http://www.ars-grin.gov/cgi-bin/npgs/html/taxon.pl?15378>.
5. Mozaffarian V. *A Dictionary of Iranian Plant Names*. Farhang Moaser Publishers, Tehran, Iran. 1998, 671 pp.
6. Delazar A, Sarker SD, Nahar L, Barzegar Jalali S, Modaresi M, Hamedeyazdan S, Babaei H, Javadzadeh Y, Asnaashari S and Bamdad Moghadam S. Rhizomes of *Eremostachys laciniata*: isolation and structure elucidation of chemical constituents and a clinical trial on inflammatory diseases. *Advanced Pharmaceutical Bulletin*, 2013; 3: 385 - 93.
7. Said O, Khalil K, Fulder S and Azaizeh H. Ethnopharmacological survey of medicinal herbs in Israel, the Golan heights and the west bank region. *Journal of Ethnopharmacol.* 2002; 83: 251 - 65.
8. Erdemoglu N, Turan NN, Cakoco I, Sener B and Aydon A. Antioxidant activities of some Lamiaceae plant extracts. *Phytotherapy Res.* 2006; 20: 9 - 13.
9. Modaresi M, Delazar A, Nazemiyeh H, Fathi-Azad F, Smith E, Rahman MM, Gibson S, Nahar R and Sarkaer S. Antibacterial iridoid glucosides from *Eremostachys laciniata*. *Phytotherapy Res.* 2009; 23: 99 - 103.
10. Nisar M, Khan S, Dar A, Rehman W, Khan R and Jan I. Antidepressant screening and flavonoids isolation from *Eremostachys laciniata* (L) Bunge. *African Journal of Biotechnol.* 2010; 10: 1696 - 99.
11. Khan S, Nisar M, Rehman W and Nasir F. Anti-inflammatory study on crude methanol extract and different fractions of *Eremostachys laciniata*. *Pharmaceutical Biol.* 2010; 48: 1115 - 18.
12. Delazar A, Habibi Asl H, Mohammadi O, Afshar FH, Nahar L, Modarresi M, Nazemiyeh H and Sarker S. Evaluation of analgesic activity of *Eremostachys laciniata* in mice. *Journal of Natural Remedies* 2009; 9: 1 - 7.
13. Azizian D and Culter DF. Anatomical, cytological and phytochemical studies on *Phlomis* L. and *Eremostachys* Bunge (Labiatae). *Botanical Journal of the Linnaean Society* 1982; 85: 249 - 81.
14. Gella EV and Vavilova NK. Monoterpene glycosides of *Eremostachys fetissovii*. *Khimicheskaya Promyshlennost Segodnya*, 1981; 3: 390 - 91.
15. Delazar A, Modarresi M, Shoeb M, Nahar L, Reid RG, Kumarasamy Y, Sarker SD. Eremostachiin: a new furanolabdane diterpene glycoside from *Eremostachys glabra*. *Natural Product Res.* 2006; 20: 167 - 72.
16. Delazar A, Shoeb M, Kumarasamy Y, Byres M, Nahar L, Modarresi M, and Sarker SD. Two bioactive ferulic acid derivatives from *Eremostachys glabra*. *DARU* 2004; 12: 49 - 53.
17. Delazar A, Gibbons S, Kumarasamy Y, Nahar L, Shoeb M and Sarker SD. Antioxidant phenylethanoid glycosides from the rhizomes of *Eremostachys glabra* (Lamiaceae). *Biochemical Systematics and Ecology* 2005; 33: 87 - 90.
18. Mughal UR, Fatima I, Malik A and Tareen RB. Loasifolin, a new flavonoid from *Eremostachys loasifolia*. *Journal of Asian Natural Products Res.* 2010; 12: 328-330.



- 19.** Calis I, Guvenc A, Armagan M, Koyuncu M, Gotfredsen CH and Jensen SR. Secondary metabolites from *Eremostachys laciniata*. *Natural Products Communications*, 2007; 3: 117-24.
- 20.** Najafpour Navaei M and Mirza M. Chemical composition of the oil of *Eremostachys laciniata* (L.) Bunge from Iran. *Flavour and Fragrance Journal*, 2006; 21: 645 - 646.
- 21.** El-Naggar LJ and Beal JL. Iridoids. A review. *Journal of Natural Products*, 1980; 43: 649 - 707.
- 22.** Dinda B, Debnath S and Harigaya Y. Naturally occurring iridoids. A review, part 1. *Chemical and Pharmaceutical Bulletin* 2007a; 55: 159 - 222.
- 23.** Dinda B, Debnath S and Harigaya Y. Naturally occurring secoiridoids and bioactivity of naturally occurring iridoids and secoiridoids. A review, part 2. *Chemical and Pharmaceutical Bulletin*, 2007b; 55: 689-728.
- 24.** Dinda B, Rou Chowdhury D and Mohanta BC. Naturally occurring iridoids, secoiridoids and their bioactivity. An updated review, part 3. *Chemical and Pharmaceutical Bulletin*, 2009; 57: 765-796.
- 25.** Dinda B, Debnath S and Banik R. Naturally occurring iridoids and secoiridoids. An updated review, part 4. *Chemical and Pharmaceutical Bulletin*, 2011; 59: 803-833.
- 26.** Rechinger KH. *Flora Iranica*. Vol. 150, Graz: Druck, 1982.
- 27.** Gutfinger T. Polyphenols in olive oils. *Journal of the American Oil Chemists Society*, 1981; 58: 966-968.
- 28.** Wagner H and Bladt S. *Plant drug analysis: A thin layer chromatography atlas*. 2<sup>nd</sup> edition. Springer, Germany. 1996, 368 pp.
- 29.** Javadi H, Hesamzadeh-Hejazi SM and Babayev MS. Comparison of karyotypic traits of *Thymus* species in Iran. *Annals of Biological Res.* 2013; 4: 199 - 208.
- 30.** Parr AJ and Bolwell GP. Phenols in the plant and in man. The potential for possible nutritional enhancement of the diet by modifying phenol content and profile. *Journal of the Science of Food and Agriculture* 2000; 80: 985 – 1012.
- 31.** Poiroux-Gonard F, Bidet LPR, Fanciullino AL, Gautier H, Lauri-Lopez F and Urban L. Health benefits of vitamins and secondary metabolites of fruits and vegetables and prospects to increase their concentrations by agronomic approaches. *Journal of Agricultural and Food Chem.* 2010; 58: 12065 - 82.
- 32.** Scalzo J, Politi A, Pellegrini N, Mezzetti B and Battino M. Plant genotype affects total antioxidant capacity and phenolic content in fruit. *Nutrition* 2005; 21: 207 - 213.
- 33.** Kwee E and Niemeyer E. Variations in phenolic composition and antioxidant properties among 15 basil (*Ocimum basilicum* L.) cultivars. *Food Chem.* 2011; 128: 1044 -50.
- 34.** Asgari Nematian M, Atri M and Nazem H. Phelavenoids diversity in *Astragalus gossypinus* (as medicinal plants) in west of Iran. *Peyk Noor Elm.* 2011; 1 (4): 50 - 61.
- 35.** Aroodi M, Ghorbanli M.L and Ahmadi-Golsefidi M. Survey of phytochemical *Stachys byzantina* C. Koch. aerial parts in North of Iran (Chaharbagh Mountain). *Journal on Plant Science Res.* 2011; 22 (2): 40 - 49.
- 36.** Hadipour A, Azizi M, Naghdi-Badi H,



Delazar A, Panahandeh J and Aroei H. Morphological diversity in some populations of *Eremostachys laciniata* Bunge. *Iranian Journal of Horticultural Sci.* 1394; 46 (3) :497 - 507.

**37.** Namjoyan F, Bazvand M and Azemi M. Antioxidant activity and Phytochemical investigation of *Onosma chlorotricum* Boiss & Noe lipophilic extract on Thin Layer

Chromatography (TLC). *Jentashapir*, 2012; 1: 55 - 60.

**38.** Hadipour A, Azizi M, Naghdi-Badi H, Delazar A, Panahandeh J and Aroei H. Karyotype study on 15 populations of *Eremostachys laciniata* Bunge in Iran. *Journal of Horticultural Science & Biotechnol.* 2016; 91(1): 55-62.